

A New Tricyclic Alkaloid from *Portulaca oleracea* L.

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A new tricyclic alkaloid named portulacatone (**1**), *i.e.*, 5,6-dihydro-8,9-dihydroxy-11*H*-pyrrolo[2,1-*b*][3]benzazepin-11-one, together with eight known compounds, methyl 4-hydroxyphenylacetate (**2**), *p*-hydroxybenzaldehyde (**3**), vanillin (**4**), protocatechualdehyde (**5**), *p*-hydroxybenzoic acid (**6**), iseluxine (**7**), oleracein E (**8**), and (+)-(*R*)-feruloyl malate (**9**) were isolated from aerial parts of *Portulaca oleracea* L. Their structures were elucidated based on spectroscopic analyses. Among them, compounds **1–7** and **9** were isolated from this medicinal plant for the first time. Compounds **1** and **7** showed dose-dependent scavenging activities against DPPH (2,2-diphenyl-1-picryl-hydrazyl) free radical, with EC_{50} values of 14.36 μM and 9.98 μM , respectively, more potent than the natural antioxidant vitamin C (EC_{50} 20.72 μM).

Introduction. – *Portulaca oleracea* L. (Purslane in English; Machixian in Chinese; Portulacacea) is an edible and medicinal plant which is widely spread throughout the world [1]. The aerial part of this plant is recorded officially in the Chinese Pharmacopoeia [2]. Pharmacological research has indicated that *P. oleracea* possesses a variety of pharmacological effects, such as anti-bacterial [3], anti-inflammatory [4], antioxidant [5], neuroprotection [6], antidiabetic [7], and antitumor [8] activities. Up to now, various kinds of constituents have been isolated from this plant, including organic acids [9], terpenoids [10], alkaloids [11–13], homoisoflavonoids [14], steroids, and phenolic compounds [15]. In our continuous search for bioactive constituents from *P. oleracea*, one new phenolic tricyclic alkaloid bearing a rare seven-membered ring, *i.e.*, 5,6-dihydro-8,9-dihydroxy-11*H*-pyrrolo[2,1-*b*][3]benzazepin-11-one (**1**), along with eight known compounds, methyl 4-hydroxyphenylacetate (**2**) [16], *p*-hydroxybenzaldehyde (**3**) [17], vanillin (**4**) [17], protocatechualdehyde (**5**) [18], *p*-hydroxybenzoic acid (**6**) [19], iseluxine (**7**) [20], oleracein E (**8**) [11], and (+)-(*R*)-feruloyl malate (**9**) [21][22] (*Fig. 1*) were isolated. Among them, compounds **1–7** and **9** were isolated from this medicinal plant for the first time. Herein, we report the structure elucidation of compound **1**, and DPPH free radical scavenging activities of **1** and **7**.

Results and Discussion. – Compound **1** was obtained as brown solid with a molecular formula as $\text{C}_{13}\text{H}_{11}\text{NO}_3$, according to the signals of m/z 230.0812 ($[M+H]^+$) and m/z 252.0632 ($[M+Na]^+$) in the HR-ESI-MS. It showed blue fluorescence under UV at 365 nm and brown-red color when sprayed with FeCl_3 solution, revealing that **1** was a phenolic compound. In the $^1\text{H-NMR}$ spectrum of compound **1** (*Table*), five unsaturated H-atoms can be observed, among which two were isolated aromatic H-atoms ($\delta(\text{H})$ 6.62 (*s*, H–C(7)), 7.46 (*s*, H–C(10))), the other three were *cis*-coupled

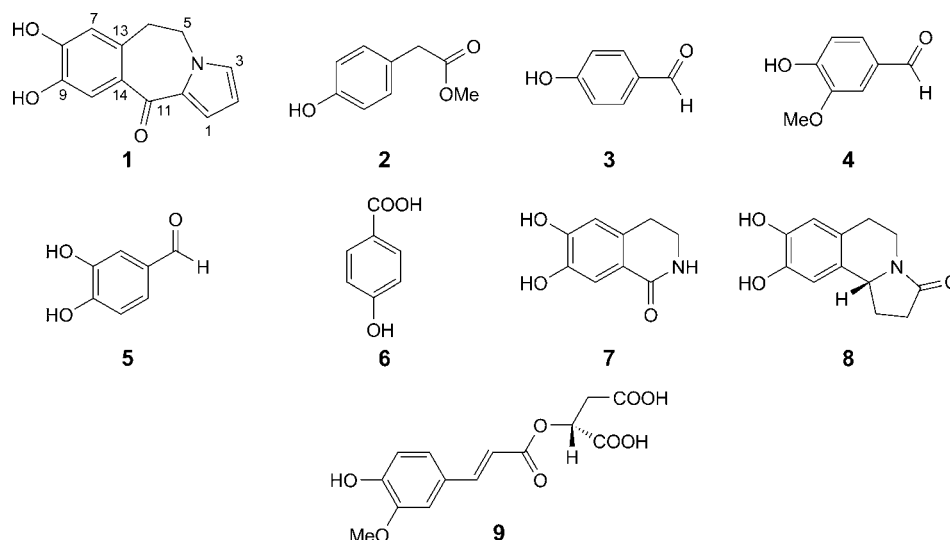


Fig. 1. Structures of compounds **1–9**, isolated from *P. oleracea*

olefinic H-atoms ($\delta(\text{H})$ 7.09 (*dd*, $J = 2.4, 1.8$, H–C(3)), 7.02 (*dd*, $J = 4.2, 1.8$, H–C(2)), 6.12 (*dd*, $J = 4.2, 2.4$, H–C(1))). In addition to the unsaturated H-atoms, the signals of a moiety linked to an N-atom were observed at $\delta(\text{H})$ 4.27 (*t*, $J = 4.8$, CH₂(5)) and 3.05 (*t*, $J = 4.8$, CH₂(6)), respectively. The ¹³C-NMR spectrum (Table) and HMQCs displayed signals of 13 C-atoms, including one C=O group at $\delta(\text{C})$ 178.9, two CH₂ groups at $\delta(\text{C})$ 50.0, 35.2, three olefinic CH groups at $\delta(\text{C})$ 109.0, 120.1, and 128.8, two aromatic CH groups at $\delta(\text{C})$ 117.0, 118.3, two O-bearing aromatic C-atoms at $\delta(\text{C})$ 150.3, and 144.6, and other three unsaturated quaternary C-atoms at $\delta(\text{C})$ 133.61, 133.63, and 127.9. The presence of a benzene ring with two OH groups located at C(8) and C(9) was confirmed by HMBCs (Fig. 2) of H–C(7)/C(8,9,14) and H–C(10)/C(8,9,13,14). In addition, the HMBCs H–C(7)/C(6), and H–C(10)/C(11) demonstrated that the benzene ring was connected with one C=O group and one CH₂ group, respectively. Furthermore, the N-connected CH₂ group at $\delta(\text{H})$ 4.27 (CH₂(5)) showed HMBCs with C(3), C(6), C(12), and C(13), whereas the three olefinic CH groups showed HMBCs with the corresponding C-atoms as indicated in Fig. 2. The HMBCs further demonstrated that the CH₂CH₂ fragment is connected to a pyrrole ring, and with a benzene ring. Therefore, a seven-membered ring was established containing C(13) and C(14) of the benzene ring, the C=O C-atom, C(12) and N from the pyrrole ring, and the CH₂CH₂ moiety. From the above analysis, the structure of compound **1** was elucidated

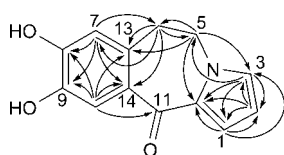


Fig. 2. Key HMBCs (H → C) of **1**

Table. ^1H - and ^{13}C -NMR Data (600 and 125 MHz, resp.; DMSO(D_6)) of Compound **1**. δ in ppm, J in Hz. Atom numbering as indicated in Fig. 1.

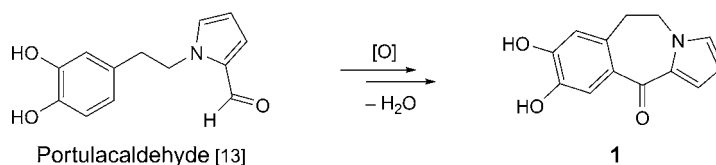
	$\delta(\text{H})$	$\delta(\text{C})$	HMBC (H \rightarrow C)
1	6.12 (<i>dd</i> , $J = 4.2, 2.4$)	109.0	2, 3, 12
2	7.02 (<i>dd</i> , $J = 4.2, 1.8$)	120.1	1, 3, 12
3	7.09 (<i>dd</i> , $J = 2.4, 1.8$)	128.8	1, 2, 5, 12
5	4.27 (<i>t</i> , $J = 4.8$)	50.0	3, 6, 12, 13
6	3.05 (<i>t</i> , $J = 4.8$)	35.2	5, 7, 13, 14
7	6.62 (<i>s</i>)	117.0	6, 8, 9, 14
8		144.6	
9		150.3	
10	7.46 (<i>s</i>)	118.3	8, 9, 11, 13, 14
11		178.9	
12		133.63	
13		133.61	
14		127.9	

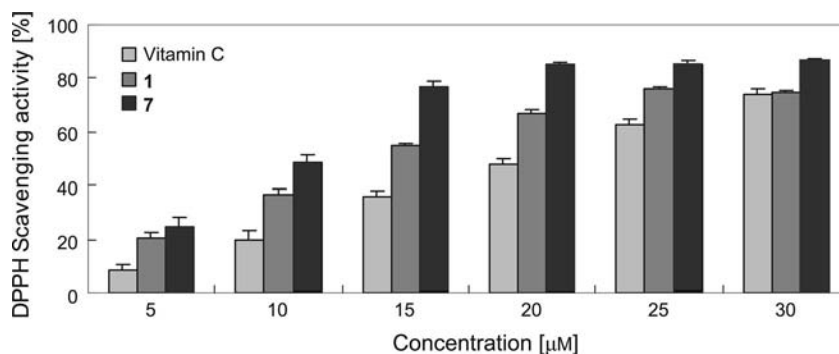
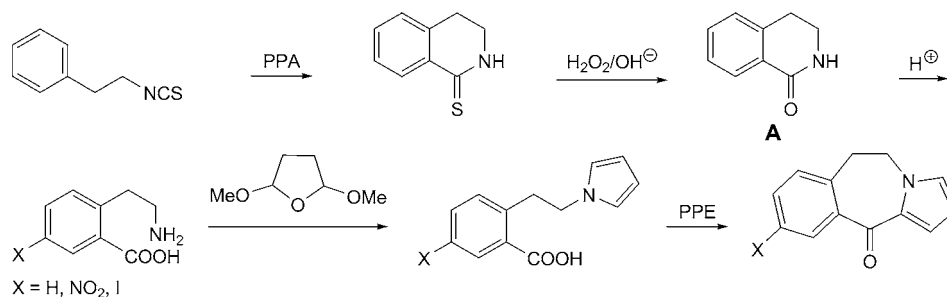
as 5,6-dihydro-8,9-dihydroxy-11*H*-pyrrolo[2,1-*b*][3]benzazepin-11-one, and named portulacatone.

In 2012, a new pyrrole alkaloid named portulacaldehyde was isolated from *P. oleracea* [13]. From a biosynthetic point of view, compound **1** can be considered as an intramolecular *Friedel–Crafts* acylation product of portulacaldehyde after oxidation to the corresponding carboxylic acid (*Scheme 1*). Moreover, the strategy for a chemical synthesis of tricyclic alkaloid derivatives of 5*H*-benzo[*d*]pyrrolo[1,2-*a*]-azepin-11(6*H*)-one was tentatively described by *Girard et al.* in 1983 [23], since these compounds were heteroanalogues of the dibenzo[*a,d*]cycloheptene ring system, a rich source of very useful drugs, particularly for treatment of diseases implicating in the central nervous system, such as cyclobenzaprine, a skeletal muscle relaxant.

Additionally, it should be noted that compound **7** was elucidated as 6,7-dihydroxy-3,4-dihydroisoquinolinone, *i.e.*, iseluxine, based on analysis of NMR spectra (^1H -, ^{13}C -, HMQC, and HMBC). This amide was firstly isolated in 2000 from the epigeal part of *Iseia luxurians* (MORIC.) O'DONELL (Convolvulaceae), a climber indigenous to the tropical Americas [20]. Interestingly, 3,4-dihydroisoquinolinone (**A**) without 6,7-dihydroxy group was an intermediate product in the chemical synthesis of substituted tricyclic alkaloid of 5*H*-benzo[*d*]pyrrolo[1,2-*a*]-azepin-11(6*H*)-one (*Scheme 2*) [23].

Compounds **1** and **7** showed dose-dependent scavenging activities against the DPPH free radical with EC_{50} values of 14.36 μM and 9.98 μM , respectively, as shown in Fig. 3, more potent than natural antioxidant vitamin C (EC_{50} 20.72 μM). Considering

Scheme 1. Possible Biosynthetic Pathway to Compound **1**

Scheme 2. 3,4-Dihydroisoquinolinone (**A**) as an Intermediate in a Synthesis of the Tricyclic Alkaloid 5H-Benzo[d]pyrrolo[1,2-a]-azepin-11(6H)-oneFig. 3. DPPH Scavenging activities of compounds **1**, **7**, and positive control vitamin C ($n=3$)

that alkaloids commonly possess a variety of biological activities, further chemical syntheses and in-depth bioactivity research on compounds **1** and **7** need to be addressed in the future.

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Experimental Part

General. TLC: Silica gel GF 254 (Qingdao Haiyang Chemical Group Co. Ltd., P. R. China); polyamide film (Taizhou Luqiao Sijia Biochemical Plastics Factory, P. R. China); visualization under UV 365 nm and 254 nm or by heating the plates sprayed with 10% H₂SO₄/EtOH, or 5% FeCl₃/EtOH, or by iodine staining. Column chromatography(CC): polyamide gel (100–200 mesh; Taizhou Luqiao Siqing biochemical Factory, P. R. China), silica gel (SiO₂, 200–300 mesh; Qingdao Haiyang Chemical Group Co. Ltd., P. R. China), MCI gel (CHP-20P, 75–150 µm, Mitsubishi Chemical Co., Japan), Sephadex LH-20 (Pharmacia Fine Chemicals, USA), and ODS-C₁₈ (75 µm, YMC Co., Japan). Semi-prep. HPLC: Shimadzu Prominence LC-20A liquid chromatography, with LC-20AT pumps, SPD-20A UV detector (Shimadzu Co., Japan), and a YMC-Pack ODS-A column (250 mm × 10 mm, 5 µm; YMC Co., Japan). Optical rotations: digital automatic polarimeter (Kernchen Co., Germany). UV Spectra: UV-2450 spectrophotometer (Shimadzu Co., Japan); λ_{max} (log ε) in nm. NMR Spectra: Agilent 600 MHz DD2;

δ in ppm rel. to Me_4Si as internal standard, J in Hz. HR-ESI-MS: *LTQ-Orbitrap* mass spectrometer (*Thermo Fisher Co.*, USA); in m/z . Microplate reader (model 680 UV, *Bio-Rad Co.*, USA) was used in the microplate assay.

Plant Material. The dried aerial parts of *P. oleracea* were purchased from *Jianlian Pharmacy* (Jinan, P.R. China) in May 2012 and were identified by Prof. *L. Xiang*, School of Pharmaceutical Sciences, Shandong University, P.R. China. A voucher specimen (No. 20120501) has been deposited with the Laboratory of Pharmacognosy, School of Pharmaceutical Sciences, Shandong University.

Extraction and Isolation. The dried sliced aerial parts (4 kg) of *P. oleracea* were refluxed with 60% EtOH (3×24 l). The combined extracts were concentrated under reduced pressure to the concentration of 0.5 g crude drug/ml (8 l). The supernatant (6 l) was subjected twice to polyamide CC (10×100 cm) eluted with gradient EtOH/ H_2O (0:100–90:10) then $\text{NH}_3 \cdot \text{H}_2\text{O}$ soln. to afford nine fractions (*Fr.* 1–9). *Fr.* 3, *Fr.* 4, and *Fr.* 6 were partitioned by AcOEt to obtain the AcOEt fractions (*Fr.* 3.1 (1.2476 g), *Fr.* 4.1 (1.7642 g), and *Fr.* 6.1 (0.873 g)), and the H_2O fractions (*Fr.* 3.2, *Fr.* 4.2, and *Fr.* 6.2). *Fr.* 6.1 was subjected to CC (*Sephadex LH-20* (4×90 cm); MeOH/ H_2O 80:20) to give 14 fractions (*Fr.* 6.1.1–6.1.14). *Fr.* 6.1.8 (89.5 mg) was then purified by CC (*Sephadex LH-20* (2×85 cm); MeOH/ H_2O 80:20) to afford three fractions *Fr.* 6.1.8.1–6.1.8.3. *Fr.* 6.1.8.2 (26 mg) was further purified by semi-prep. HPLC (MeOH/0.1% HCOOH 40:60) to obtain compound **1** (3.1 mg). *Fr.* 3.1 was subjected to CC (*Sephadex LH-20* (4×90 cm); MeOH/ H_2O 80:20) to give *Fr.* 3.1.1–3.1.18. *Fr.* 3.1.11 (217 mg) was then subjected to CC (SiO_2 , PE/AcOEt 8:2 to 5:5) to yield six fractions. *Fr.* 3.1.11.1–3.1.11.6. *Fr.* 3.1.11.1 was further purified by semi-prep. HPLC (MeOH/0.1% HCOOH 40:60) to obtain compound **2** (2 mg). *Fr.* 3.1.11.2 was further purified by semi-prep. HPLC (MeOH/0.1% HCOOH 32:68) to obtain compounds **3** (1.6 mg) and **4** (3.5 mg). *Fr.* 3.1.12 (88 mg) was repurified by CC (*Sephadex LH-20*; MeOH/ H_2O 80:20; followed by *MCI* (2.5×22 cm); MeOH/ H_2O 0:100–80:20) and semi-prep. HPLC (MeOH/0.1% HCOOH 20:80) to provide compound **7** (5 mg). *Fr.* 4.1 was subjected to CC (*Sephadex LH-20* (4×90 cm); MeOH/ H_2O 80:20) to give 14 fractions: *Fr.* 4.1.1–4.1.14. *Fr.* 4.1.6 (237 mg) was then purified by CC (*Sephadex LH-20* (2×85 cm); MeOH/ H_2O 80:20) to afford five fractions: *Fr.* 4.1.6.1–4.1.6.5. *Fr.* 4.1.6.4 (70 mg) was further purified by CC (*Sephadex LH-20* (2×85 cm); MeOH/ H_2O 80:20) to yield four fractions *Fr.* 4.1.6.4.1–4.1.6.4.4. *Fr.* 4.1.6.4.3 (26 mg) was then purified by semi-prep. HPLC (MeOH/0.1% HCOOH 37:63) to obtain compound **5** (2.5 mg). *Fr.* 4.1.6.4.4 (12.4 mg) was purified by semi-prep. HPLC (MeOH/0.1% HCOOH 28:72) to obtain compound **6** (6 mg). Compound **8** (15 mg) was precipitated and purified from *Fr.* 4.1.7. *Fr.* 9 (3.0405 g) was first subjected to CC (*Sephadex LH-20* (6×59 cm), MeOH/ H_2O 80:20) to obtain *Fr.* 9.1–9.9. *Fr.* 9.5 was then separated by CC (*Sephadex LH-20* (6×59 cm), MeOH/ H_2O 80:20) to give *Fr.* 9.5.1–9.5.17. *Fr.* 9.5.6 was subjected to CC (*ODS-C₁₈* (3.5×8.5 cm), EtOH/ H_2O 0:10 to 10:0) to yield *Fr.* 9.5.6.1–9.5.6.11. *Fr.* 9.5.6.1 was repurified by CC (*ODS-C₁₈* (2×25 cm), EtOH/ H_2O 0:10 to 10:0) to afford *Fr.* 9.5.6.1.1–9.5.6.1.3. *Fr.* 9.5.6.1.1 was last purified by semi-prep. HPLC (MeOH/ H_2O 43:57) to provide compound **9** (5 mg) with $[\alpha]_D^{20}$ value of $+7.40$ ($c = 0.1$, MeOH).

Portulacatone (=5,6-Dihydro-8,9-dihydroxy-11H-pyrrolo[2,1-b][3]benzazepin-11-one; **1**). Brown solid. UV (MeOH): 346.1 (4.07), 317.6 (3.99), 246.9 (3.89). ^1H - and ^{13}C -NMR: Table. HR-ESI-MS: 230.0812 ($[M+H]^+$, $\text{C}_{13}\text{H}_{12}\text{NO}_3^+$; calc. 230.0817), 252.0632 ($[M+Na]^+$, $\text{C}_{13}\text{H}_{11}\text{NNaO}_3^+$; calc. 252.0637).

DPPH Scavenging Activity Assay. The experiment was carried out according to the reported method [24] with a little modification. Briefly, 25 μl sample soln. (dissolved in MeOH) and 200 μl DPPH (100 μM , dissolved in MeOH) were mixed in 96-well plates. After shaking for 30 min at r.t. in the dark, the absorbance was measured at 490 nm by microplate reader and recorded as $A_{\text{sample+DPPH}}$. The absorbance of 25 μl sample mixed with 200 μl MeOH was recorded as A_{sample} . The absorbance of 25 μl MeOH mixed with 200 μl DPPH was recorded as A_{DPPH} , with the absorbance of 225 μl MeOH as the blank and recorded as A_{blank} . Each sample was examined for three times. The percentage inhibition was calculated according to the following equation:

$$\text{DPPH scavenging activity (\%)} = [1 - (A_{\text{sample+DPPH}} - A_{\text{sample}}) / (A_{\text{DPPH}} - A_{\text{blank}})] \times 100$$

The EC_{50} value was obtained by interpolation from linear regression analysis, which represented the concentration of sample that decreased the DPPH free radical by 50%.

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